



# Exogenous progestogens differentially alter gene expression of immature cumulus–oocyte complexes in sheep



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## ABSTRACT

This study evaluated the role of progesterone (P<sub>4</sub>) and medroxyprogesterone acetate (MAP) on the molecular status of immature cumulus–oocyte complexes (COCs) and the implications for oocyte quality in sheep. The number of viable COCs per ewe and the rate of COCs screened for developmental competence by brilliant cresyl blue positive (BCB<sup>+</sup>) were similar ( $P > 0.05$ ), respectively, across treatments (P<sub>4</sub>:  $7.7 \pm 0.7$  and  $4.7 \pm 1.2$ ; MAP:  $5.7 \pm 1.0$  and  $3.5 \pm 2.3$ ; and control:  $5.7 \pm 1.1$  and  $3.6 \pm 2.4$ ). The COCs' gene expression was altered by exogenous progestogens compared with the control group: markers of steroidogenic pathway (FSH receptor [FSHr], LH receptor [LHr], and estradiol receptor  $\alpha$ ) and of quality (zygote arrest 1, growth differentiation factor 9, and B-cell lymphoma 2) were in abundance in P<sub>4</sub> ( $P < 0.05$ ). In addition, reelin protein (RELN) was downregulated, and Bcl-2 was upregulated in MAP ( $P < 0.05$ ). In the P<sub>4</sub> vs MAP comparison, FSHr, LHr, and RELN genes were upregulated ( $P < 0.05$ ) in the P<sub>4</sub> group. In conclusion, P<sub>4</sub> and MAP promoted dissimilar effects on transcriptome profiling of immature BCB-selected COCs, possibly due to the differences in the chemical structure of progestogens and concentrations of serum P<sub>4</sub>. Exogenous P<sub>4</sub> impacted positively on the profile of genes related to oocyte quality.

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## 1. Introduction

Hormonal manipulation of ovarian function using the association of gonadotropins and progestogens is widely applied in assisted reproductive technologies to improve oocyte recovery in live females of monovular species [1–3]. Stimulatory protocols with FSH increase follicular population [4,5] and alter global gene expression in oocyte and cumulus cells [6–8]. Progestogen support during gonadotropin treatments is used to inhibit LH pulse frequency, allowing regression of dominant follicle(s) by atresia, and thus avoiding ovulation [2,9].

Progestogens ( $P_4$ ) are natural or synthetic steroids that exert  $P_4$ -like activity. Progesterone is the only natural progestogen and is produced primarily by corpus luteum after ovulation. Progesterone analogs, such as medroxyprogesterone acetate (MAP), simulate the therapeutic effects of  $P_4$ , despite some differences in chemical structure. These differences result in different actions at the cellular level [10,11]. Progesterone has an important role in mammalian oocyte maturation, but its impact on oocyte quality has not yet been properly clarified [12]. In addition, no information is available about the effect of  $P_4$  vs MAP during antral follicular growth on the gene expression of immature cumulus-oocyte complexes (COCs).

Recent evidence using intravaginal  $P_4$  administration during FSH stimulation of the first wave of the estrous cycle showed that sheep embryo yield is improved in *in vivo* [13] and *in vitro* [14] production systems. The authors demonstrated that this improvement was associated with a positive effect of  $P_4$  on oocyte developmental competence [14]. In contrast, Bartlewski et al [15] evaluated the use of  $P_4$  vs MAP during a conventional superovulation protocol and observed no difference in terms of number of ovulations and embryos recovered. Conversely, some studies reported that exogenous progestogens had a deleterious effect on *in vitro* embryo production [1], mainly in long-term based treatments [3]. Nonetheless, there is a gap in the literature about the impact of exogenous progestogens at molecular level and the consequences to immature oocytes. The expression profile of genes related to the acquisition of oocyte competence, such as growth factors, maternal effect, steroidogenic pathway, apoptosis, and regulatory proteins, among others, could help in the development of more effective synchronization and/or stimulation protocols for use in assisted reproduction *in vivo* and *in vitro*.

Oocyte quality is related to developmental competence, as oocytes sustain profound changes at structural, molecular, and biochemical levels, including maternal zygotic transition. This capability is acquired during the follicular growth, mainly the final phase. However, reaching full growth does not ensure that an oocyte is capable of undergoing all the steps of embryonic development to term [16,17]. Nevertheless, studies have demonstrated that oocytes, which have already finished the exponential growth phase, screened by brilliant cresyl blue (BCB), are more competent and show a greater blastocyst rate. The BCB test measures the activity of glucose-6-phosphate dehydrogenase, which has low activity in fully grown oocytes (BCB<sup>+</sup>) and high activity in growing oocyte (BCB<sup>-</sup>) [18,19]. Thus, we tested the hypothesis that  $P_4$  and MAP promote dissimilar effects on the molecular status of COCs impacting oocyte quality. The aim of the present study was to evaluate the effect of  $P_4$  and MAP on the gene expression profile of immature COCs and the implication of this on oocyte quality prediction in sheep.

## 2. Materials and methods

### 2.1. Local conditions and animal care

The study was performed at the *Unidade de Pesquisa Experimental em Caprinos e Ovinos*, at *Universidade Federal*

*Fluminense*, in Cachoeiras de Macacu, Rio de Janeiro, Brazil (22°27' S, 43°39' W) in October of 2017 (nonbreeding season). All procedures were approved by the local Ethical Committee for Animal Use of the University (protocol #721/2015) and were conducted under the ethical principles of the Brazilian Society of Animal Experimentation. Thirty multiparous Santa Inês ewes (mean  $\pm$  SD: 3.9  $\pm$  1.0 yr old, 51  $\pm$  5.9 kg of BW, and 3.2  $\pm$  0.6 of BCS/scale 0–5) were used; the animals were clinically healthy, fed on chopped elephant grass (*Pennisetum purpureum*) and 200 g per animal of concentrate (12% crude protein) twice daily, and receiving water and mineralized salt *ad libitum*.

### 2.2. Experimental design

Thirty estrus-synchronized ewes were submitted to FSH ovarian stimulation at ovulation (Day 0), in the first follicular wave. At the moment of the first FSH administration, ewes were allocated into 3 experimental groups (n = 10) and received an intravaginal device containing either  $P_4$  or MAP, or no device (control), as shown in Figure 1. Follicular aspiration was performed on Day 2 by laparoscopic ovum pick-up (LOPU), and recovered COCs were morphologically graded and tested with BCB. Selected BCB<sup>+</sup> COCs were used for gene expression analysis. Blood samples were collected from the jugular vein using vacutainer tubes to determine serum  $P_4$  concentrations from Day 1 to Day 2.

### 2.3. Estrus synchronization

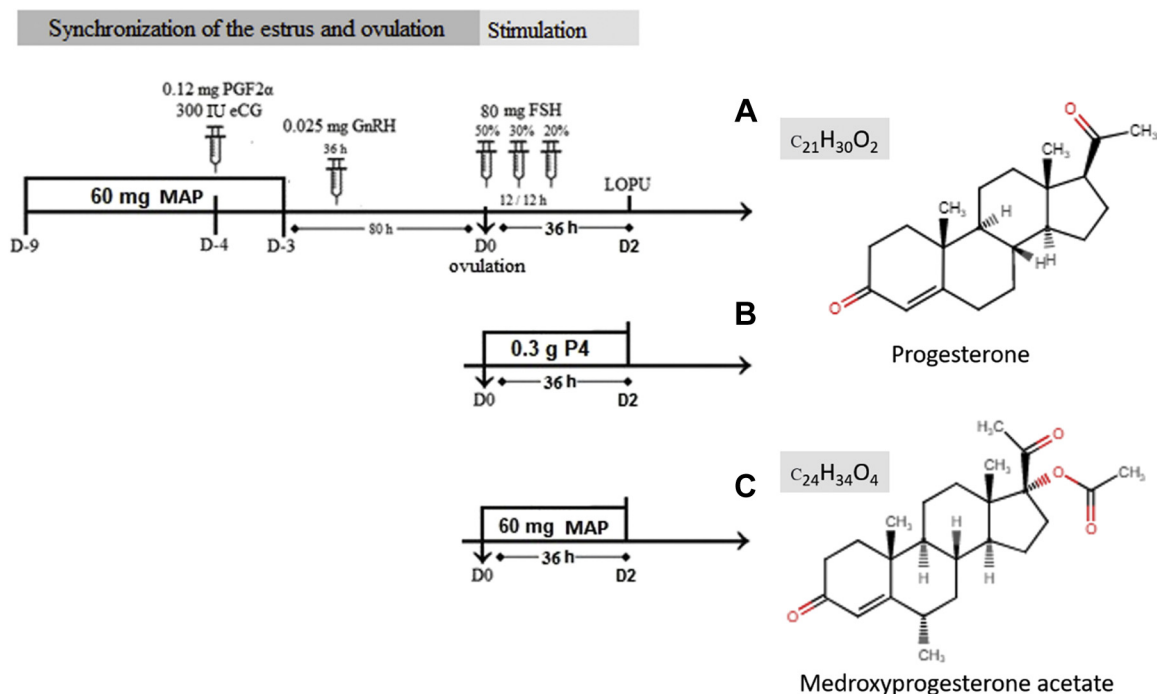
All ewes received an estrus synchronization treatment using a short-term protocol described by Balara et al [22]. Briefly, intravaginal sponges containing 60 mg MAP (Progespon; Schering Plough Animal Health, SP, Brazil) were applied for 6 d. One day before sponge removal, 300 IU eCG (Novormon 5000; MSD Animal Health, SP, Brazil) and 0.12 mg cloprostenol sodium (Estron, Tecnopec, São Paulo, Brazil) were administered intramuscularly (i.m.). At 36 h after sponge removal, 0.025 mg lecorelin (Gestran Plus; Tecnopec, SP, Brazil) was given i.m.

### 2.4. Progestogen sources during ovarian stimulation

At 80 h after sponge withdrawal (defined as Day 0, ie, soon after ovulation) 80 mg of FSH (Folltropin-V; Vetoquinol, Paris, France) were administered in 3 injections (50%, 30%, and 20%) every 12 h to stimulate the first follicular wave of the estrous cycle. At the first FSH dose, the ewes received either an intravaginal device containing 0.33 mg of  $P_4$  (CIDR, Eazi-Breed; Zoetis;  $P_4$  group) or an intravaginal sponge containing 60 mg MAP, whereas the control group did not receive any progestogen device and remained untreated during the early luteal phase (ie, endogenous  $P_4$ ).

### 2.5. Serum $P_4$ determinations

Blood samples were collected and centrifuged (2,600  $\times$  g), and serum was stored at  $-20^\circ\text{C}$  until  $P_4$  determination, which was performed by a solid-phase radioimmunoassay using a commercial kit (ImmuChem; MP Biomedicals, Santa



**Fig. 1.** Representative scheme of the experimental design. At 80 h after sponge removal of the estrus synchronization treatment (Day 0), ovarian stimulation was performed with 80 mg of FSH in 3 decreasing doses every 12 h. At first FSH dose, the females were allocated to one of 3 experimental groups: (A) No progestogen support was applied in the control group (exposed to endogenous progesterone); (B) progesterone ( $P_4$ ) was administered by intravaginal devices (CIDR-G); and (C) medroxyprogesterone acetate (MAP) was administered by intravaginal devices (sponges). (A) and (B) were exposed to endogenous and endogenous plus exogenous  $P_4$  (bioidentical, same chemical structure), respectively, while (C) was exposed to MAP synthetic progestogen analog (with a different chemical structure). Source for molecule structure [20,21].

Ana, CA, USA). The sensitivity and intra-assay coefficient were 0.05 ng/mL and 11%, respectively. All data were within the maximum and minimum point of the curve.

## 2.6. COCs selection and BCB test

COCs were recovered by LOPU, as previously described [8]. COCs were isolated under a stereomicroscope, and their quality was graded based on a visual assessment of morphology (number of cellular layers and cytoplasmic uniformity), with Grade I/II being good, III acceptable, and IV poor [23]. To predict developmental competence, viable COCs (GI, GII, and GIII) from each treatment were washed once and exposed to 26  $\mu$ M BCB (B5388, Sigma) diluted in DMPBS supplemented with 10% BSA (A9647, Sigma) and 0.2 mM pyruvate (P4562, Sigma), for 60 min on a hot plate at 37°C and ambient atmosphere. Then, COCs were washed twice, placed in polyvinyl alcohol (P8136; Sigma) for 5 min, and classified according to oocyte cytoplasm staining as BCB<sup>+</sup> (blue cytoplasm) and BCB<sup>-</sup> (colorless cytoplasm). BCB<sup>+</sup> COCs from each treatment were recovered with a minimal amount of medium and transferred to DNase/RNase-free cryotubes, subjected to snap freezing, and stored in liquid nitrogen canisters for later gene expression analysis.

## 2.7. Gene expression analysis

Samples were analyzed by quantitative PCR (qPCR) after reverse transcription [24]. Total RNA was extracted from 3

pools of 5 COCs BCB<sup>+</sup> per group using the RNeasyMicro Kit (Qiagen Inc, Valencia, EUA) according to the manufacturer's instructions and treated with DNase for 15 min to prevent DNA contamination. Elution was performed with 14  $\mu$ L of RNAase free water, and the RNA quantification of each pool was performed using 1  $\mu$ L of sample on a spectrophotometer (NanoDrop 2000, Wilmington, DE, USA). For reverse transcription, using the SuperScript III first-strand synthesis Supermix (Invitrogen, Carlsbad, CA, USA), the same RNA concentration was used for all samples. The reverse transcription reaction was prepared by mixing oligo (dT) 20 primers, dNTP mixture, superscript III RT, RNase OUT, MgCl<sub>2</sub>, RT buffer, and RNA sample in a final volume of 20  $\mu$ L. The mixtures were first incubated at 65°C for 5 min and then for 50°C for 50 min. The reaction was terminated at 85°C for 5 min and then chilled on ice. After that, RNase H was added to the samples and incubated at 37°C for 20 min.

Relative quantification was performed in triplicate using qPCR (ABI Prism 7300 Sequence Detection Systems, Foster City, CA, USA). Reactions (20  $\mu$ L total volume) were prepared using a mixture of SYBR green kit (10  $\mu$ L; Power SYBR Green, Applied Biosystems), 0.1  $\mu$ M primers (Table 1), nuclease-free water, and reverse-transcribed cDNA (1  $\mu$ L). Negative controls, comprising the PCR reaction mixture without nucleic acids, were also run with each group of samples. Template cDNAs were denatured at 95°C for 10 min, and all genes were amplified by 40 cycles of a thermal cycling programmed of 95°C for 15 s, 60°C for 15 s and 60°C for 30 s. Fluorescence data were acquired during the extension steps. After each

**Table 1**

Sequences of the specific primers used in the analysis of gene expression in immature brilliant cresyl blue–positive cumulus-oocyte complexes (BCB<sup>+</sup>) recovered by laparoscopy after different ovarian stimulation treatments in Santa Ines ewes.

Gene	Sequence of primers (5'- 3')	Annealing temperature (°C)	Size (bp)	Reference
GAPDH	GGGAAA TCG TGC GTG ACA TTA AG TGTTGT GCGTAAGGTCITTTG	60	273	Bebbere et al 2008
ZAR1	CAC TGC AAG GAC TGC AAT ATC CAG GTG ATA TCC TCC ACT C	60	137	Bebbere et al 2008
MATER	CAG CCT CCA GGA GTT CTT TG GAC AGC CTA GGA GGG TTT CC	59	212	Bebbere et al 2008
GDF9	CAG ACG CCA CCT CTA CAA CA CAG GAA AGG GAA AAG AAA TGG	58	198	Bebbere et al 2008
BMP15	GGG TTC TAC GAC TCC GCT TC GGT TAC TTT CAG GCC CAT CAT	59	273	Bebbere et al 2008
BAX	CCT GGG ATC TTG AAA CTC TCC TT CTG AGC CAG GCT GAA ATC AAA A	60	566	Chakravarthi et al 2017
Bcl-2	GCC GAG TGA GCA GGA AGA C GTT AGC CAG TGC TTG CTG AGA	60	214	Chakravarthi et al 2017
ER $\alpha$	GAA TCT GCC AAG GAG ACT CG CCT GAC AGC TCT TCC TCC TG	60	187	Hogg et al 2011
LHr	TCC GAA AGC TTC CAG ATG TT GAA ATC AGC GTT GTC CCA TT	60	199	Hogg et al 2011
FSHr	TAA GCA CTT GCC AGC TGT TC CTC ATC GAG TTG GGT TCC AT	60	196	Hogg et al 2011
STAR	GCA TCC TCA AAG ACC AGG AG CTT GAC ACT GGG GTT CCA CT	60	194	Hogg et al 2011
RELN	CAGCCAAAGGACTTCAACCA CGAGTGCTACTAGCACAACC	60	161	NM_001306121.1
LRP8	ACACCTGTGGCTGTGTTC CCAGAGGCAGGAAAGACCTG	60	161	NM_001097565.1

Abbreviations: BAX, BCL2-associated X protein; Bcl-2, B-cell lymphoma 2; BMP15, bone morphogenetic protein 15; ER $\alpha$ , estrogen receptor; FSHr, FSH receptor; GAPDH, glyceraldehyde3-phosphate dehydrogenase; GDF9, growth differentiation factor 9; LHr, LH receptor; LRP-8, LDL receptor–related protein 8; MATER, maternal antigen that embryo requires; RELN, reelin; STAR, steroidogenic acute regulatory protein; ZAR1, zygote arrest 1.

PCR run, a melting curve analysis was performed to confirm that a single specific product was generated. Primer efficiency was calculated using LinRegPCR software [25] for each reaction. The primer efficiency average was 1.95; 1.96; 1.92; 1.89; 1.91; 1.93; 1.91; 1.91; 1.91; 1.93; and 1.98 to ZAR1, MATER, BMP15, BAX, Bcl-2, ER $\alpha$ , LHr, FSHr, STAR, RELN, and GAPDH, respectively. Relative quantification was performed by the comparative Ct method ( $2^{-\Delta\Delta Ct}$ ) using the REST 2008 software [26,27]. The expression of each target gene was normalized using GAPDH. The groups were compared as Control vs MAP, control vs P<sub>4</sub>, and MAP vs P<sub>4</sub>. Fold changes are relative to the calibrator (=1), which was either the control (Control vs MAP and control vs P<sub>4</sub>) or MAP (MAP vs P<sub>4</sub>) data.

## 2.8. Statistical analysis

Variables with normal distribution were analyzed by ANOVA, and the significance of differences between mean values was determined using Tukey's test. Data presented as a frequency or percentage (ie, proportion of BCB<sup>+</sup> COCs) were analyzed by chi-square test. Statistical analyses were performed using a statistical analysis system program (SAEG 9.0; Universidade Federal de Viçosa). Differences were considered significant when  $P < 0.05$ . Data are given as the mean  $\pm$  SEM.

## 3. Results

### 3.1. COCs recovery and gene expression

There was no effect of progestogen source on the total number of viable or BCB<sup>+</sup> COCs, as shown in Table 2. Gene

expression was altered by exogenous progestogens compared with the control (Fig. 2A, B): Steroidogenic pathway receptors (FSHr, LHr, and ER $\alpha$ ), as well as markers of oocyte quality (ZAR1, GDF9, and Bcl-2) genes were in abundance in P<sub>4</sub> ( $P < 0.05$ ). Conversely, RELN was down-regulated, and Bcl-2 was upregulated in MAP ( $P < 0.05$ ). In the comparison between the different devices, FSHr, LHr, and RELN genes were upregulated ( $P < 0.05$ ) in P<sub>4</sub> compared with the MAP group.

### 3.2. Serum P<sub>4</sub> concentrations

All ewes from the 3 experimental groups had non-detectable serum P<sub>4</sub> concentrations 1 d before (Day -1) and on the day that treatments began (Day 0, at first FSH administration). Progesterone concentrations were affected by day and type of device ( $P < 0.05$ ; Fig. 3), this being greater on Day 1 and Day 2 in P<sub>4</sub> group ( $P < 0.05$ ), compared with the MAP and control groups. The mean concentrations on Day 2 were  $1.0 \pm 1.0$ ,  $0.6 \pm 0.3$ , and  $3.7 \pm 0.7$  ng/mL ( $P < 0.05$ ) in the control, MAP, and P<sub>4</sub> group, respectively.

## 4. Discussion

This study provides new information about the impact of exogenous progestogens on the gene expression profile of immature COCs in the sheep model. The results demonstrate that P<sub>4</sub> and MAP cause dissimilar effects on gene expression in fully grown COCs (BCB<sup>+</sup>). This effect probably occurred due to the variations in the chemical

**Table 2**

Effect of different progestogen treatments during FSH stimulation in the first follicular wave, on the quality (morphology and BCB test) of immature cumulus–oocyte complexes (COCs) per ewe recovered by laparoscopy in Santa Ines ewes (mean  $\pm$  SE).

Group	COCs selection/ewe			
	Viable COCs <sup>a</sup>	Bad COCs <sup>a</sup>	BCB <sup>+b</sup>	BCB <sup>-b</sup>
P <sub>4</sub> <sup>c</sup>	7.7 $\pm$ 0.7	0.5 $\pm$ 0.7	4.7 $\pm$ 1.2	3.2 $\pm$ 1.1
MAP <sup>c</sup>	5.7 $\pm$ 1.0	0.5 $\pm$ 0.7	3.5 $\pm$ 2.3	2.2 $\pm$ 1.5
Control <sup>c</sup>	5.7 $\pm$ 1.1	0.2 $\pm$ 0.4	3.6 $\pm$ 2.4	2.1 $\pm$ 1.7

Abbreviation: BCB, brilliant cresyl blue positive.

n = 10 ewes per treatment; ( $P > 0.05$ ).

<sup>a</sup> Viable COCs: grading according to the cumulus cellular layers and cytoplasmic homogeneity, as good (G1, multilayered compacted cumulus and homogeneous ooplasm; G2, one to 3 layers and homogeneous ooplasm) and acceptable (G3, one incomplete layer or denuded but homogeneous ooplasm); \*Bad COCs: G4 (shapeless, expanded cumulus, and degenerated) were discarded.

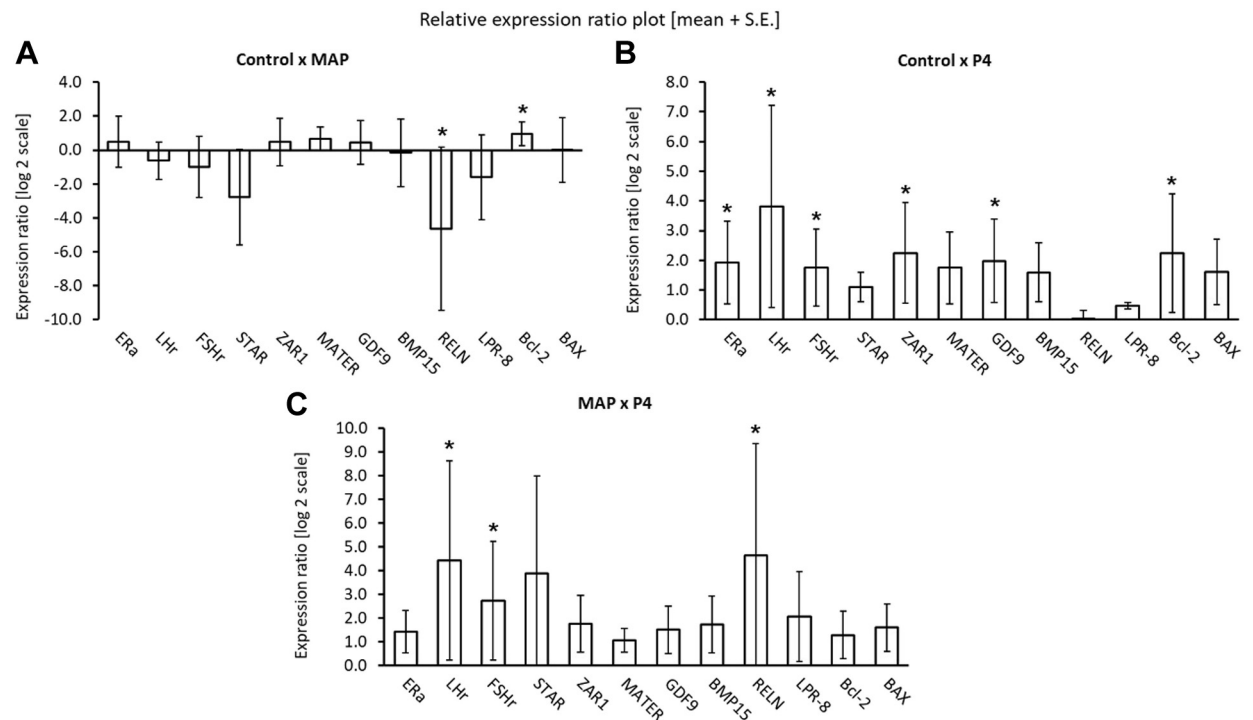
<sup>b</sup> BCB<sup>+</sup>: competent fully grown COCs; BCB<sup>-</sup>: noncompetent growing COCs.

<sup>c</sup> P<sub>4</sub>: Ewes received CIDR containing 0.33 mg of progesterone (P<sub>4</sub>); MAP: ewes received sponge containing 60 mg of medroxyprogesterone acetate; Control: ewes did not receive any progestogen device.

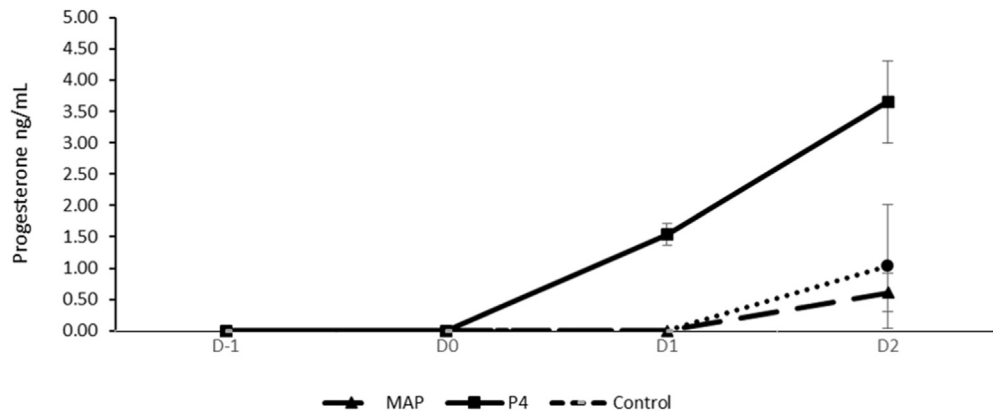
structure of these progestogens, resulting in different activity and inducing different transcriptome profiling.

BCB test showed that oocyte exponential growth was not influenced by progestogen because there was no difference in the number of BCB<sup>+</sup> or BCB<sup>-</sup> COCs across the P<sub>4</sub>, MAP, and control groups. In other words, all the groups had oocytes of similar competence prediction. The higher developmental competence of the BCB<sup>+</sup> COCs compared with the BCB<sup>-</sup> COCs may be related to the favorable conditions of the follicular development during the final

phases of folliculogenesis and has already been demonstrated in sheep [19,28] and cows [18,29]. Therefore, only BCB<sup>+</sup> COCs were used in the present study, resulting in more homogeneous samples, which enabled the attribution of differences in gene expression induced by the treatments. In addition, our recent data [5] showed no differences in follicular population and COC morphological features (GI, GII, GIII, and GIV) across the groups. Thus, FSH applied for ovarian stimulation appears to be the responsible to determine the number, growth, and morphological



**Fig. 2.** Relative gene expression in immature cumulus–oocyte complexes (COCs), brilliant cresyl blue–positive (BCB<sup>+</sup>) derived from nonprogestogen-treated (control) or progestogen-treated (MAP or P<sub>4</sub>) ewes during ovarian stimulation of the follicular first wave. The groups were compared as (A) Control vs MAP, (B) Control vs P<sub>4</sub>, and (C) MAP vs P<sub>4</sub>. Fold changes are relative to the calibrator (=1), which was the control data in (A and B) and MAP data in (C). Data show means  $\pm$  SE \* $P < 0.05$  compared with reference gene. GAPDH, glyceraldehyde3–phosphate dehydrogenase; ER $\alpha$ , estrogen receptor; LHr, LH receptor; FSHr, FSH receptor; STAR, steroidogenic acute regulatory protein; ZAR1, zygote arrest 1; MATER, maternal antigen that embryo requires; GDF9, growth differentiation factor 9; BMP15, bone morphogenetic protein 15; RELN, reelin; LPR-8, LDL receptor–related protein 8; Bcl-2, B-cell lymphoma 2; BAX, BCL2-associated X protein.



**Fig. 3.** Serum progesterone (P4) concentrations in Santa Ines ewes treated with progesterone (P4 group), medroxyprogesterone acetate (MAP group), and nontreated ewes (control group) during the early luteal phase. Day 0 was defined at 80 h after sponge removal in an estrus synchronization treatment.

quality of the COCs, regardless of the progestogen used. Although progestogens had no effect on ovarian dynamics [5] and COCs morphology, P<sub>4</sub> and MAP promoted different effects on gene expression of COCs (BCB<sup>+</sup>), confirming our hypothesis.

As expected, the P<sub>4</sub> treatment promoted greater P<sub>4</sub> serum concentration in comparison with MAP and control groups. Low serum P<sub>4</sub> in MAP and control groups was a consequence of the recent formation of the corpus luteum. These data are in agreement with Cuadro et al [13], who reported higher P<sub>4</sub> serum concentration in ewes receiving a P<sub>4</sub> implant than ewes in the early luteal phase (control) during the FSH stimulation of the first follicular wave. Similarly, Bartlewski et al [15] observed a greater P<sub>4</sub> concentration in exogenous P<sub>4</sub>-treated ewes than in MAP-treated ewes during a superovulation protocol. It is well known that MAP serum concentration is not detected by the RIA kit used to measure P<sub>4</sub>. For MAP detection, it is necessary to use a specific antiserum [30,31]. We speculate that this may occur because of the divergence in the chemical structure between MAP and P<sub>4</sub>. The MAP molecule presents a methyl (CH<sub>3</sub>) radical at C6 position and a methyl acetate (C<sub>2</sub>H<sub>3</sub>O<sub>2</sub>) radical at C17 position (Fig. 1), whereas in the P<sub>4</sub> molecule, this radical is absent. The exogenous P<sub>4</sub> is bioidentical to the endogenous (luteal) hormone [11].

The abundance of messenger RNA (mRNA) encoding proteins expressed in the oocyte (ie, *MATER*, *ZAR1*, *BMP15*, and *GDF9*), cumulus (ie, *RELN*, *LRP8*, *FSHr*, *LHr*, *ERα*, and *STAR*) or both (ie, *BAX* and *Bcl-2*) were evaluated in BCB<sup>+</sup> COCs. In the P<sub>4</sub> group, 6 genes were upregulated (*ZAR1*, *GDF9*, *Bcl2*, *FSHr*, *LHr*, and *ERα*), and in the MAP group, 2 genes were affected (*RELN* was downregulated and *Bcl2* upregulated), both compared with the control (calibrator). However, comparing P<sub>4</sub> vs MAP (calibrator), 3 genes were upregulated (*RELN*, *FSHr*, and *LHr*) in the P<sub>4</sub> group.

The difference in *RELN* abundance (downregulated in MAP compared with control and upregulated in P<sub>4</sub> compared with MAP) was not related to serum P<sub>4</sub> concentration because *RELN* expression was no different between P<sub>4</sub> and control. The contrast in *RELN* mRNA expression may be due to the different chemical structure

of the progestogens. We presume that P<sub>4</sub> and MAP pharmacokinetics activate distinct pathways, despite the similar therapeutic effect (pharmacodynamics). *RELN* expression is maximal in the theca cells of dominant follicles and binding *LRP8* in granulosa cells by paracrine action, activating the downstream signaling pathways [32], which regulate the final follicle growth stage [7]. Yang et al [33] reported *RELN* involvement with protein kinases activity and suggested it plays a role in progestogenic pathways. Conversely, Fayad et al [32] have demonstrated that *RELN* and its ligand, *LRP8*, are predominantly expressed in the dominant follicles and similarly downregulated during the ovulatory process and luteinization. Thus, we considered the *RELN* abundance as an important marker of oocyte competence acquisition.

We associate the gene expression profile observed in the P<sub>4</sub> group to the rise of serum P<sub>4</sub> concentrations. Recently, Menchaca et al [14] demonstrated that high serum P<sub>4</sub> induced by a P<sub>4</sub> intravaginal device enhances oocyte developmental competence by increasing the in vitro cleavage rate and embryo development in sheep (compared with nontreated ewes). Cuadro et al [13] have reported similar effects in the in vivo embryo production system in sheep, which both did and did not receive exogenous P<sub>4</sub> during the growing phase of the follicular wave. The authors found an improvement in the percentage of fertilized oocytes after insemination, a greater number of transferable embryos per donor, and better quality of the collected embryos. Otherwise, when P<sub>4</sub> priming was compared with MAP treatment, Bartlewski et al [15] reported no differences in terms of the number of corpora lutea and embryos recovered in superovulated ewes.

In support of the upregulation of *ZAR1* in the P<sub>4</sub>-treated group in relation to the control group, we considered this a good indication of quality improvement; moreover, it may corroborate the findings of Menchaca et al [14] and Cuadro et al [13], already mentioned. *ZAR1* is a good predictor of developmental competence, mainly in immature oocytes. *ZAR1* mRNA plays a role in early embryo development until the zygotic genome activation [34,35].

*FSH* levels directly influence *FSHr*, *LHr*, and *ERα* [8,36] by downstream activation of a *FSHr*/adenylylase/cAMP/

PKA cascade to trigger estradiol ( $E_2$ ) production [37–39]. An interaction between  $P_4$  and FSH has already been evidenced [14,40], and we suggest that this interaction could be a function of the  $P_4$  molecular structure; it may be that the methyl acetate radical on MAP molecules may prevent this interaction. Bartlewski et al [15] observed longer FSH-metabolic clearance rates in the  $P_4$ -treated than the MPA-treated ewes. Based on this finding, and those reported by Wei et al [36] and Knecht et al [41], we speculate that the longer FSH bioavailability in  $P_4$ -treated ewes possibly maintained the stimulus of downstream FSH-induced pathways, enabling the expression of these receptors, including LHR, through raising of cAMP, despite  $P_4$  inhibition of LH release pulsatile frequency by feedback mechanism. The upregulation of these receptors (*FSHR*, *LHR*, and *ER $\alpha$* ) may promote better conditions for oocyte to respond to the in vitro maturation conditions regarding the role of hormonal regulation during the maturation process.

The greater serum  $P_4$  also affected *GDF9* expression, probably by extending FSH bioavailability in the  $P_4$ -treated group. Exogenous FSH affects *GDF9*, as we have previously demonstrated [8]. *GDF9* stimulates granulosa cell proliferation and acts in the regulatory process during final follicle growth, before the LH surge before ovulation; cumulus cells require *GDF9* to support the metabolic cascades and steroid production [42,43]. Hence, the  $P_4$ -treated group seems to promote more appropriate COC gene modulation.

Apoptosis proteins regulate cell death and are good markers of cell viability. The antiapoptotic *Bcl2* gene was upregulated in both the  $P_4$ - and MAP-treated groups in the present study. Similar results were obtained by Thammissiri et al [3] in a short-term MAP-treated protocol. These evidence indicate the positive effect of exogenous progestogens applied during ovarian stimulation on the apoptosis modulation cascade.

Nilsen and Brinton [44] demonstrated the divergent impact of  $P_4$  and MAP on the mitogen-activated protein kinase (MAPK) pathway for the mechanism of neuroprotection in neuron culture, which reinforces our hypothesis about both progestogens activating different downstream pathways, despite their similar therapeutic effects. Exogenous support of  $P_4$  seems to exert a beneficial effect on oocyte quality, based on the profile of gene expression (compared with the MAP-treated and control groups). On the other hand, MAP had a negative effect, mainly because of the downregulation of *RELN*, since this extracellular matrix glycoprotein is involved in 47 distinct cell functions, including MAPK pathway [33], which is necessary for germinal vesicle breakdown and oocyte maturation [45]. However, to confirm this deleterious effect of MAP, future research is necessary to evaluate in vitro development and molecular features of the embryos, such as polyspermy regulation, cryopreservation, and/or implantation.

In conclusion, despite  $P_4$  and MAP promoted obtaining COCs with similar quality related to morphological features and BCB test, they induced dissimilar effects on the molecular status of immature COCs in sheep. Exogenous  $P_4$  appears to influence indirectly the oocyte competence through its positive impact on the COC gene

expression (*ZAR1*, *GDF9*, *Bcl2*, *FSHR*, *LHR*, and *ER $\alpha$* ), probably because of its chemical structure (identical to the  $P_4$  endogenous) and greater serum  $P_4$  concentration, which induce this difference in its biological activity. However, the exact mechanism by which it occurs still needs to be elucidated.

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## References

- [1] Berlinguer F, Gonzalez-Bulnes A, Succu S, Leoni G, Mossa F, Bebbere D, et al. Effects of progestagens on follicular growth and oocyte developmental competence in FSH-treated ewes. *Domest Anim Endocrinol* 2007;32:303–14.
- [2] Kuang Y, Chen Q, Fu Y, Wang Y, Hong Q, Lyu Q, et al. Medroxyprogesterone acetate is an effective oral alternative for preventing premature luteinizing hormone surges in women undergoing controlled ovarian hyperstimulation for in vitro fertilization. *Fertil Steril* 2015;104:62–70.e3.
- [3] Thammissiri J, Navanukraw C, Uriyapongson S, Khanthusaseng V, Kamollirt C. Expression of cumulus-oocyte complex genes and embryonic development in goats subjected to progestogen-based estrus synchronization. *Theriogenology* 2015;86:612–8.
- [4] Mendes CS, Barbosa LP, de Araújo ML, Gomes Pinheiro EE, Lents MP, da Silva Amorim L, et al. Follicular response and oocyte production following variations in ovarian stimulation in goats. *Theriogenology* 2018;108:88–96.
- [5] Bragança GM, Souza-Fabjan JMG, Ribeiro LS, Brair VL, Côrtes LR, Souza CV, et al. Exogenous progestogen does not affect first-wave follicle populations and oocyte quality during ovarian stimulation with FSH in sheep. *Domest Anim Endocrinol* 2019;106369.
- [6] Dias FCF, Khan MIR, Sirard MA, Adams GP, Singh J. Differential gene expression of granulosa cells after ovarian superstimulation in beef cattle. *Reproduction* 2013;146:181–91.
- [7] Nivet A-L, Vigneault C, Blondin P, Sirard M-A. Changes in granulosa cells' gene expression associated with increased oocyte competence in bovine. *Reproduction* 2013;145:555–65.
- [8] Bragança GM, Batista RITP, Souza-Fabjan JMG, Alfradique VAP, Arashiro EKN, Cosentino IO, et al. Dose and administration protocol for FSH used for ovarian stimulation affect gene expression in sheep cumulus-oocyte complexes. *Reprod Fertil Dev* 2018;30:1234–44.
- [9] Greyling JPC, Van Der Nest M. Synchronization of oestrus in goats: dose effect of progestagen. *Small Rumin Res* 2000;36:201–7.
- [10] Lieberman A, Curtis L. In defense of a review. *Altern Ther Health Med* 2017;23:24–32.
- [11] L'Hermite M. Bioidentical menopausal hormone therapy: registered hormones (non-oral estradiol  $\pm$  progesterone) are optimal. *Climacteric* 2017;20:331–8.
- [12] Lonergan P. Influence of progesterone on oocyte quality and embryo development in cows. *Theriogenology* 2011;76:1594–601.
- [13] Cuadro F, dos Santos-Neto PC, Pinczak A, Barrera N, Crispo M, Menchaca A. Serum progesterone concentrations during FSH superstimulation of the first follicular wave affect embryo production in sheep. *Anim Reprod Sci* 2018;196:205–10.
- [14] Menchaca A, Cuadro F, dos Santos-Neto PC, Bosolasco D, Barrera N, de Brun V, et al. Oocyte developmental competence is improved by relatively greater circulating progesterone concentrations during preovulatory follicular growth. *Anim Reprod Sci* 2018;195:321–8.
- [15] Bartlewski PM, Seaton P, Szpila P, Oliveira MEF, Murawski M, Schwarz T, et al. Comparison of the effects of pretreatment with Veramix sponge (medroxyprogesterone acetate) or CIDR (natural progesterone) in combination with an injection of estradiol-17 $\beta$  on ovarian activity, endocrine profiles, and embryo yields in cyclic ewes superovu. *Theriogenology* 2015;84:1225–37.

- [16] Sirard MA, Richard F, Blondin P, Robert C. Contribution of the oocyte to embryo quality. *Theriogenology* 2006;65:126–36.
- [17] Sirard MA. Follicle environment and quality of in vitro matured oocytes. *J Assist Reprod Genet* 2011;28:483–8.
- [18] Torner H, Ghanem N, Ambros C, Hölker M, Tomek W, Phatsara C, et al. Molecular and subcellular characterisation of oocytes screened for their developmental competence based on glucose-6-phosphate dehydrogenase activity. *Reproduction* 2008;135:197–212.
- [19] Mohammadi-Sangcheshmeh A, Veshkini A, Hajarizadeh A, Jamshidi-Adegani F, Zhandi M, Abazari-Kia AH, et al. Association of glucose-6-phosphate dehydrogenase activity with oocyte cytoplasmic lipid content, developmental competence, and expression of candidate genes in a sheep model. *J Assist Reprod Genet* 2014;31:1089–98.
- [20] PubChem Database. National Center for Biotechnology Information. Progesterone, CID=5994. <https://pubchem.ncbi.nlm.nih.gov/compound/Progesterone>. [Accessed March 2020].
- [21] PubChem Database. National Center for Biotechnology Information. Medroxyprogesterone acetate, CID=6279. <https://pubchem.ncbi.nlm.nih.gov/compound/Medroxyprogesterone-acetate>. [Accessed March 2020].
- [22] Balaro MFA, Fonseca JF, Barbosa TGB, Souza-Fabjan JMG, Figueira LM, Teixeira TA, et al. Potential role for GnRH in the synchronization of follicular emergence before the superovulatory Day 0 protocol. *Domest Anim Endocrinol* 2016;54:10–4.
- [23] Souza-Fabjan JMG, Pereira AF, Melo CHS, Sanchez DJD, Oba E, Mermillod P, et al. Assessment of the reproductive parameters, laparoscopic oocyte recovery and the first embryos produced in vitro from endangered Canindé goats (*Capra hircus*). *Reprod Biol* 2013;13:325–32.
- [24] Batista RITP, Raposo NRB, Campos-Junior PHA, Pereira MM, Camargo LSA, Carvalho BC, et al. Trans-10, cis-12 conjugated linoleic acid reduces neutral lipid content and may affect cryotolerance of in vitro-produced crossbred bovine embryos. *J Anim Sci Biotechnol* 2014;5:1–8.
- [25] Ramakers C, Ruijter JM, Lekanne Deprez RH, Moorman AFM. Assumption-free analysis of quantitative real-time polymerase chain reaction (PCR) data. *Neurosci Lett* 2003;339:62–6.
- [26] Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the  $2^{-\Delta\Delta C_T}$  Method. *Methods* 2001;408:402–8.
- [27] Pfaffl MW. Relative expression software tool (REST(C)) for group-wise comparison and statistical analysis of relative expression results in real-time PCR. *Nucleic Acids Res* 2002;30:e36.
- [28] Mohammadi-Sangcheshmeh A, Soleimani M, Deldar H, Salehi M, Soudi S, Hashemi SM, et al. Prediction of oocyte developmental competence in ovine using glucose-6-phosphate dehydrogenase (G6PDH) activity determined at retrieval time. *J Assist Reprod Genet* 2012;29:153–8.
- [29] Alm H, Torner H, Lörke B, Viergutz T, Ghoneim IM, Kanitz W. Bovine blastocyst development rate in vitro is influenced by selection of oocytes by brilliant cresyl blue staining before IVM as indicator for glucose-6-phosphate dehydrogenase activity. *Theriogenology* 2005;63:2194–205.
- [30] Ortiz A, Hiroi M, Stanczyk FZ, Goebelsmann U, Mishell DR. Serum medroxyprogesterone acetate (MPA) concentrations and ovarian function following intramuscular injection of depo-mpa. *J Clin Endocrinol Metab* 1977;44:32–9.
- [31] Shrimanker K, Saxena BN, Fotherby K. A radioimmunoassay for serum medroxyprogesterone acetate. *J Steroid Biochem* 1978;9:359–63.
- [32] Fayat T, Lefebvre R, Nimpf J, Silversides DW, Lussier JG. Low-density lipoprotein receptor-related protein 8 (LRP8) is upregulated in granulosa cells of bovine dominant follicle: molecular characterization and spatio-temporal expression studies. *Biol Reprod* 2007;76:466–75.
- [33] Yang F, Wang M, Zhang B, Xiang W, Zhang K, Chu M, et al. Identification of new progesterone-associated networks in mammalian ovulation using bioinformatics. *BMC Syst Biol* 2018;6:1–12.
- [34] Bebbere D, Bogliolo L, Ariu F, Fois S, Leoni GG, Tore S, et al. Expression pattern of zygote arrest 1 (ZAR1), maternal antigen that embryo requires (MATER), growth differentiation factor 9 (GDF9) and bone morphogenetic protein 15 (BMP15) genes in ovine oocytes and in vitro-produced preimplantation embryos. *Reprod Fertil Dev* 2008;20:908.
- [35] Uzbekova S, Roy-Sabau M, Dalbiès-Tran R, Perreau C, Papillier P, Mompert F, et al. Zygote arrest 1 gene in pig, cattle and human: evidence of different transcript variants in male and female germ cells. *Reprod Biol Endocrinol* 2006;4:1–14.
- [36] Wei S, Shen X, Gong Z, Deng Y, Lai L, Liang H. FSHR and LHR expression and signaling as well as maturation and apoptosis of cumulus-oocyte complexes following treatment with FSH receptor binding inhibitor in sheep. *Cell Physiol Biochem* 2017;43:660–9.
- [37] Millier SG, Whitelaw PF, Smyth CD. Follicular oestrogen synthesis: the “two-cell, two-gonadotrophin” model revisited. *Mol Cell Endocrinol* 1994;100:51–4.
- [38] Silva JM, Hamel M, Sahmi M, Price CA. Control of oestradiol secretion and of cytochrome P450 aromatase messenger ribonucleic acid accumulation by FSH involves different intracellular pathways in oestrogenic bovine granulosa cells in vitro. *Reproduction* 2006;132:909–17.
- [39] Ulloa-Aguirre A, Reiter E, Crepieux P. FSH receptor signaling: complexity of interactions and signal diversity. *Endocrinology* 2018;159:3020–35.
- [40] Baby TE, Bartlewski PM. Progesterone as the driving regulatory force behind serum FSH concentrations and antral follicular development in cycling ewes. *Reprod Fertil Dev* 2011;23:303–10.
- [41] Knecht M, Darbon JM, Ranta T, Baukal AJ, Catt KJ. Estrogens enhance the adenosine 3',5'-monophosphate-mediated induction of follicle-stimulating hormone and luteinizing hormone receptors in rat granulosa cells. *Endocrinology* 1984;115:41–9.
- [42] Otsuka F, McTavish K, Shimasaki S. Integral role of GDF-9 and BMP-15 in ovarian function. *Bone* 2011;78:9–21.
- [43] Shimasaki S, Moore M, Otsuka F, Erickson G. The bone morphogenetic protein system in mammalian reproduction. *Endocr Rev* 2004;25:72–101.
- [44] Nilsen J, Brinton RD. Divergent impact of progesterone and medroxyprogesterone acetate (Provera) on nuclear mitogen-activated protein kinase signaling. *Proc Natl Acad Sci* 2003;100:10506–11.
- [45] Silvestre F, Boni R, Fissore RA, Tosti E. Ca<sup>2+</sup> signaling during maturation of cumulus-oocyte complex in mammals. *Mol Reprod Dev* 2011;78:744–56.