Exogenous progestogens differentially alter gene expression of immature cumulus–oocyte complexes in sheep


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Article info
This study evaluated the role of progesterone (P4) and medroxyprogesterone acetate (MAP) on the molecular status of immature cumulus–oocyte complexes (COCs) and the implications for oocyte quality in sheep. The number of viable COCs per ewe and the rate of COCs screened for developmental competence by brilliant cresyl blue positive (BCB+) were similar (P > 0.05), respectively, across treatments (P4: 7.7 ± 0.7 and 4.7 ± 1.2; MAP: 5.7 ± 1.0 and 3.5 ± 2.3; and control: 5.7 ± 1.1 and 3.6 ± 2.4). The COCs’ gene expression was altered by exogenous progestogens compared with the control group: markers of steroidogenic pathway (FSH receptor [FSHr], LH receptor [LHr], and estradiol receptor α) and of quality (zygote arrest 1, growth differentiation factor 9, and B-cell lymphoma 2) were in abundance in P4 (P < 0.05). In addition, reelin protein (RELN) was downregulated, and Bcl-2 was upregulated in MAP (P < 0.05). In the P4 vs MAP comparison, FSHr, LHr, and RELN genes were upregulated (P < 0.05) in the P4 group. In conclusion, P4 and MAP promoted dissimilar effects on transcriptome profiling of immature BCB-selected COCs, possibly due to the differences in the chemical structure of progestogens and concentrations of serum P4. Exogenous P4 impacted positively on the profile of genes related to oocyte quality.

1. Introduction
Hormonal manipulation of ovarian function using the association of gonadotropins and progestogens is widely applied in assisted reproductive technologies to improve oocyte recovery in live females of monovular species [1–3]. Stimulatory protocols with FSH increase follicular population [4,5] and alter global gene expression in oocyte and cumulus cells [6–8]. Progestogen support during gonadotropin treatments is used to inhibit LH pulse frequency, allowing regression of dominant follicle(s) by atresia, and thus avoiding ovulation [2,9].

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Progestogens (P₄) are natural or synthetic steroids that exert P₄-like activity. Progesterone is the only natural progestogen and is produced primarily by corpus luteum after ovulation. Progesterone analogs, such as medroxyprogesterone acetate (MAP), simulate the therapeutic effects of P₄, despite some differences in chemical structure. These differences result in different actions at the cellular level [10,11]. Progesterone has an important role in mammalian oocyte maturation, but its impact on oocyte quality has not yet been properly clarified [12]. In addition, no information is available about the effect of P₄ vs MAP during antral follicular growth on the gene expression of immature cumulus–oocyte complexes (COCs).

Recent evidence using intravaginal P₄ administration during FSH stimulation of the first wave of the estrous cycle showed that sheep embryo yield is improved in vivo [13] and in vitro [14] production systems. The authors demonstrated that this improvement was associated with a positive effect of P₄ on oocyte developmental competence [14]. In contrast, Bartlewski et al. [15] evaluated the use of P₄ vs MAP during a conventional superovulation protocol and observed no difference in terms of number of ovarulations and embryos recovered. Conversely, some studies reported that exogenous progestogens had a deleterious effect on oocyte quality has not yet been properly clarified [12]. In addition, no information is available about the effect of P₄ vs MAP during antral follicular growth on the gene expression of immature cumulus–oocyte complexes (COCs).

2. Materials and methods

2.1. Local conditions and animal care

The study was performed at the Unidade de Pesquisa Experimental em Caprinos e Ovinos, at Universidade Federal Fluminense, in Cachoeiras de Macacu, Rio de Janeiro, Brazil (22°27′ S, 43°39′ W) in October of 2017 (nonbreeding season). All procedures were approved by the local Ethical Committee for Animal Use of the University (protocol #721/2015) and were conducted under the ethical principles of the Brazilian Society of Animal Experimentation. Thirty multiparous Santa Inês ewes (mean ± SD: 3.9 ± 1.0 yr old, 51 ± 5.9 kg of BW, and 3.2 ± 0.6 of BCS/scale 0–5) were used; the animals were clinically healthy, fed on chopped elephant grass (Pennisetum purpureum) and 200 g per animal of concentrate (12% crude protein) twice daily, and receiving water and mineralized salt ad libitum.

2.2. Experimental design

Thirty estrus-synchronized ewes were submitted to FSH ovarian stimulation at ovulation (Day 0), in the first follicular wave. At the moment of the first FSH administration, ewes were allocated into 3 experimental groups (n = 10) and received an intravaginal device containing either P₄ or MAP, or no device (control), as shown in Figure 1. Follicular aspiration was performed on Day 2 by laparoscopic ovum pick-up (LOPU), and recovered COCs were morphologically graded and tested with BCB. Selected BCB COCs were used for gene expression analysis. Blood samples were collected from the jugular vein using vacutainer tubes to determine serum P₄ concentrations from Day 1 to Day 2.

2.3. Estrus synchronization

All ewes received an estrus synchronization treatment using a short-term protocol described by Balaro et al. [22]. Briefly, intravaginal sponges containing 60 mg MAP (Progesteron; Schering Plough Animal Health, SP, Brazil) were applied for 6 d. One day before sponge removal, 300 IU eCG (Novormon 5000; MSD Animal Health, SP, Brazil) and 0.12 mg cloprostenol sodium (Estron, Tecnopec, São Paulo, Brazil) were administered intramuscularly (i.m.). At 36 h after sponge removal, 0.025 mg lecirelin (Gestran Plus; Tecnopec, SP, Brazil) was given i.m.

2.4. Progestogen sources during ovarian stimulation

At 80 h after sponge withdrawal (defined as Day 0, ie, soon after ovulation) 80 mg of FSH (Folltropin-V; Vetquinoil, Paris, France) were administered in 3 injections (50%, 30%, and 20%) every 12 h to stimulate the first follicular wave of the estrous cycle. At the first FSH dose, the ewes received either an intravaginal device containing 0.33 mg of P₄ (CIDR, Eazi-Breed; Zoetis; P₄ group) or an intravaginal sponge containing 60 mg MAP, whereas the control group did not receive any progestogen device and remained untreated during the early luteal phase (ie, endogenous P₄).

2.5. Serum P₄ determinations

Blood samples were collected and centrifuged (2,600 × g), and serum was stored at −20°C until P₄ determination, which was performed by a solid-phase radioimmunoassay using a commercial kit (ImmuChem; MP Biomedicals, Santa
Ana, CA, USA). The sensitivity and intra-assay coefficient were 0.05 ng/mL and 11%, respectively. All data were within the maximum and minimum point of the curve.

2.6. COCs selection and BCB test

COCs were recovered by LOPU, as previously described [8]. COCs were isolated under a stereomicroscope, and their quality was graded based on a visual assessment of morphology (number of cellular layers and cytoplasmic uniformity), with Grade I/II being good, III acceptable, and IV poor [23]. To predict developmental competence, viable COCs (GI, GII, and GIII) from each treatment were washed once and exposed to 26 mM BCB (B5388, Sigma) diluted in DMPBS supplemented with 10% BSA (A9647, Sigma) and 0.2 mM pyruvate (P4562, Sigma), for 60 min on a hot plate at 37°C and ambient atmosphere. Then, COCs were washed twice, placed in polyvinyl alcohol (P8136; Sigma) for 5 min, and classified according to oocyte cytoplasm staining as BCB⁺ (blue cytoplasm) and BCB⁻ (colorless cytoplasm). BCB⁺ COCs from each treatment were recovered with a minimal amount of medium and transferred to DNAse/RNAsase-free cryotubes, subjected to snap freezing, and stored in liquid nitrogen canisters for later gene expression analysis.

2.7. Gene expression analysis

Samples were analyzed by quantitative PCR (qPCR) after reverse transcription [24]. Total RNA was extracted from 3 pools of 5 COCs BCB⁺ per group using the RNeasy Micro Kit (Qiagen Inc, Valencia, EUA) according to the manufacturer’s instructions and treated with DNase for 15 min to prevent DNA contamination. Elution was performed with 14 μL of RNAase free water, and the RNA quantification of each pool was performed using 1 μL of sample on a spectrophotometer (NanoDrop 2000, Wilmington, DE, USA). For reverse transcription, using the SuperScript III first-strand synthesis Supermix (Invitrogen, Carlsbad, CA, USA), the same RNA concentration was used for all samples. The reverse transcription reaction was prepared by mixing oligo (dT) 20 primers, dNTP mixture, superscript III RT, RNase OUT, MgCl2, RT buffer, and RNA sample in a final volume of 20 μL. The reaction was terminated at 85°C for 5 min and then chilled on ice. After that, RNase H was added to the samples and incubated at 37°C for 20 min.

Relative quantification was performed in triplicate using qPCR (ABI Prism 7300 Sequence Detection Systems, Foster City, CA, USA). Reactions (20 μL total volume) were prepared using a mixture of SYBR green kit (10 μL; Power SYBR Green, Applied Biosystems), 0.1 μM primers (Table 1), nuclease-free water, and reverse-transcribed cDNA (1 μL). Negative controls, comprising the PCR reaction mixture without nucleic acids, were also run with each group of samples. Template cDNAs were denatured at 95°C for 10 min, and all genes were amplified by 40 cycles of a thermal cycling programmed of 95°C for 15 s, 60°C for 15 s and 60°C for 30 s. Fluorescence data were acquired during the extension steps. After each
Abbreviations: BAX, B-cell lymphoma 2-associated X protein; Bcl-2, B-cell lymphoma 2; BMP15, bone morphogenetic protein 15; ER, estrogen receptor; FSHr, FSH receptor; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; GDF9, growth differentiation factor 9; LHr, LH receptor; LRP-8, LDL receptor-related protein 8; MATER, maternal antigen that embryo requires; RELN, reelin; STAR, steroidogenic acute regulatory protein; ZAR1, zygote arrest 1.

**Table 1**
Sequences of the specific primers used in the analysis of gene expression in immature brilliant cresyl blue-positive cumulus-oocyte complexes (BCB⁺) recovered by laparoscopy after different ovarian stimulation treatments in Santa Ines ewes.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Sequence of primers (5’-3’)</th>
<th>Annealing temperature (°C)</th>
<th>Size (bp)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>GAPDH</td>
<td>GGGAAA TCG TGC GTG ACA TTA AG TGTGTT GGGGTAGAACCTTTGT</td>
<td>60</td>
<td>273</td>
<td>Bebbere et al. 2008</td>
</tr>
<tr>
<td>ZAR1</td>
<td>CAC TGC AAG GAC TGC AAT ATC CAG GTG ATA TCC TCC ACT C</td>
<td>60</td>
<td>137</td>
<td>Bebbere et al. 2008</td>
</tr>
<tr>
<td>MATER</td>
<td>CAG CCT CCA GGA GTC CTT TG</td>
<td>59</td>
<td>212</td>
<td>Bebbere et al. 2008</td>
</tr>
<tr>
<td>GDP9</td>
<td>CAG ACG CAA CCT CTA CAA CA</td>
<td>58</td>
<td>198</td>
<td>Bebbere et al. 2008</td>
</tr>
<tr>
<td>BMP15</td>
<td>GGG TTC TAC GAC TCC GCT TC CAG TAC TTT CAG GCC CAT CAT</td>
<td>59</td>
<td>273</td>
<td>Bebbere et al. 2008</td>
</tr>
<tr>
<td>BAX</td>
<td>CCT GGG ATC TTG AAA CCT TCC TT CTT AGC CAG GCT GAA ATC AAA A</td>
<td>60</td>
<td>566</td>
<td>Chakravarthi et al. 2017</td>
</tr>
<tr>
<td>Bcl-2</td>
<td>GCC GAG TGA GCA GGA ACA C</td>
<td>60</td>
<td>214</td>
<td>Chakravarthi et al. 2017</td>
</tr>
<tr>
<td>ERx</td>
<td>GAA TCT GCC AGG AGG ACT CG</td>
<td>60</td>
<td>187</td>
<td>Hogg et al. 2011</td>
</tr>
<tr>
<td>LHr</td>
<td>TCC GAA ATC TCC CAG ATG TT CAG GAC TCC TCC TG</td>
<td>60</td>
<td>199</td>
<td>Hogg et al. 2011</td>
</tr>
<tr>
<td>FSHr</td>
<td>TAA GCA GTC GCC AGG TGT TC CTC AT CAG GTC CCA TT</td>
<td>60</td>
<td>196</td>
<td>Hogg et al. 2011</td>
</tr>
<tr>
<td>RELN</td>
<td>CAGCCAAAACCTTCCACCCA CGAGTCCTATACGCAACACC</td>
<td>60</td>
<td>161</td>
<td>NM_001306121.1</td>
</tr>
<tr>
<td>LRP8</td>
<td>ACACTCCTGTCCTGTTGTC AACAAGACACG</td>
<td>60</td>
<td>161</td>
<td>NM_001097565.1</td>
</tr>
</tbody>
</table>

**Table 2**
Genes expression was altered by exogenous progestogens compared with the control (Fig. 2A, B): Steroidogenic pathway receptors (FSHr, LHr, and Erα), as well as markers of oocyte quality (ZAR1, GDF9, and Bcl-2) genes were in abundance in P4 (P < 0.05). Conversely, RELN was down-regulated, and Bcl-2 was upregulated in MAP (P < 0.05). In the comparison between the different devices, FSHr, LHr, and RELN genes were upregulated (P < 0.05) in P4 compared with the MAP group.

2.8 Statistical analysis

Variables with normal distribution were analyzed by ANOVA, and the significance of differences between mean values was determined using Tukey’s test. Data presented as a frequency or percentage (ie, proportion of BCB⁺ COCs) were analyzed by chi-square test. Statistical analyses were performed using a statistical analysis system program (SAEG 9.0: Universidade Federal de Viçosa). Differences were considered significant when P < 0.05. Data are given as the mean ± SEM.

3. Results

3.1 COCs recovery and gene expression

There was no effect of progestogen source on the total number of viable or BCB⁺ COCs, as shown in Table 2. Gene expression was altered by exogenous progestogens compared with the control (Fig. 2A, B): Steroidogenic pathway receptors (FSHr, LHr, and Erα), as well as markers of oocyte quality (ZAR1, GDF9, and Bcl-2) genes were in abundance in P4 (P < 0.05). Conversely, RELN was down-regulated, and Bcl-2 was upregulated in MAP (P < 0.05). In the comparison between the different devices, FSHr, LHr, and RELN genes were upregulated (P < 0.05) in P4 compared with the MAP group.

3.2 Serum P₄ concentrations

All ewes from the 3 experimental groups had non-detectable serum P₄ concentrations 1 d before (Day −1) and on the day that treatments began (Day 0, at first FSH administration). Progesterone concentrations were affected by day and type of device (P < 0.05; Fig. 3), this being greater on Day 1 and Day 2 in P₄ group (P < 0.05), compared with the MAP and control groups. The mean concentrations on Day 2 were 1.0 ± 1.0, 0.6 ± 0.3, and 3.7 ± 0.7 ng/mL (P < 0.05) in the control, MAP, and P₄ group, respectively.

4. Discussion

This study provides new information about the impact of exogenous progestogens on the gene expression profile of immature COCs in the sheep model. The results demonstrate that P₄ and MAP cause dissimilar effects on gene expression in fully grown COCs (BCB⁺). This effect probably occurred due to the variations in the chemical
structure of these progestogens, resulting in different activity and inducing different transcriptome profiling.

BCB test showed that oocyte exponential growth was not influenced by progestogen because there was no difference in the number of BCB⁺ or BCB⁻ COCs across the P₄, MAP, and control groups. In other words, all the groups had oocytes of similar competence prediction. The higher developmental competence of the BCB⁺ COCs compared with the BCB⁻ COCs may be related to the favorable conditions of the follicular development during the final phases of folliculogenesis and has already been demonstrated in sheep [19,28] and cows [18,29]. Therefore, only BCB⁺ COCs were used in the present study, resulting in more homogeneous samples, which enabled the attribution of differences in gene expression induced by the treatments. In addition, our recent data [5] showed no differences in follicular population and COC morphological features (GI, GII, GIII, and GIV) across the groups. Thus, FSH applied for ovarian stimulation appears to be the responsible to determine the number, growth, and morphological

Table 2
Effect of different progestogen treatments during FSH stimulation in the first follicular wave, on the quality (morphology and BCB test) of immature cumulus–oocyte complexes (COCs) per ewe recovered by laparoscopy in Santa Ines ewes (mean ± SE).

<table>
<thead>
<tr>
<th>Group</th>
<th>COCs selection/ewe</th>
<th>Viable COCsᵃ</th>
<th>Bad COCsᵇ</th>
<th>BCB⁺ᵇ</th>
<th>BCB⁻ᵇ</th>
</tr>
</thead>
<tbody>
<tr>
<td>P₄</td>
<td></td>
<td>7.7 ± 0.7</td>
<td>0.5 ± 0.7</td>
<td>4.7 ± 1.2</td>
<td>3.2 ± 1.1</td>
</tr>
<tr>
<td>MAP</td>
<td></td>
<td>5.7 ± 1.0</td>
<td>0.5 ± 0.7</td>
<td>3.5 ± 2.3</td>
<td>2.2 ± 1.5</td>
</tr>
<tr>
<td>Control</td>
<td></td>
<td>5.7 ± 1.1</td>
<td>0.2 ± 0.4</td>
<td>3.6 ± 2.4</td>
<td>2.1 ± 1.7</td>
</tr>
</tbody>
</table>

Abbreviation: BCB, brilliant cresyl blue positive.

n = 10 ewes per treatment; (P > 0.05).

ᵃ Viable COCs: grading according to the cumulus cellular layers and cytoplasmic homogeneity, as good (G1, multilayered compacted cumulus and homogeneous ooplasm; G2, one to 3 layers and homogeneous ooplasm) and acceptable (G3, one incomplete layer or denuded but homogeneous ooplasm);

ᵇ Bad COCs: G4 (shapeless, expanded cumulus, and degenerated) were discarded.

ᵇ BCB⁺: competent fully grown COCs; BCB⁻: noncompetent growing COCs.

ᶜ P₄: Ewes received CIDR containing 0.33 mg of progesterone (P₄); MAP: ewes received sponge containing 60 mg of medroxyprogesterone acetate; Control: ewes did not receive any progestogen device.

Fig. 2. Relative gene expression in immature cumulus–oocyte complexes (COCs), brilliant cresyl blue-positive (BCB⁺) derived from nonprogestogen-treated (control) or progestogen-treated (MAP or P₄) ewes during ovarian stimulation of the follicular first wave. The groups were compared as (A) Control vs MAP, (B) Control vs P₄, and (C) MAP vs P₄. Fold changes are relative to the calibrator (=1), which was the control data in (A and B) and MAP data in (C). Data show means ± SE *P < 0.05 compared with reference gene. GAPDH, glyceraldehyde3-phosphate dehydrogenase; ERα, estrogen receptor; LHr, LH receptor; FSHr, FSH receptor; STAR, steroidogenic acute regulatory protein; ZAR1, zygote arrest 1; MATER, maternal antigen that embryo requires; GDF9, growth differentiation factor 9; BMP15, bone morphogenetic protein 15; RELN, reelin; LRP-8, LDL receptor–related protein 8; Bcl-2, B-cell lymphoma 2; BAX, BCL2-associated X protein.
As expected, the P₄ treatment promoted greater P₄ serum concentration in comparison with MAP and control groups. Low serum P₄ in MAP and control groups was a consequence of the recent formation of the corpus luteum. These data are in agreement with Cuadro et al. [13], who reported higher P₄ serum concentration in ewes receiving a P₄ implant than ewes in the early luteal phase (control) during the FSH stimulation of the first follicular wave. Similarly, Bartlewski et al. [15] observed a greater P₄ concentration in exogenous P₄-treated ewes than in MAP-treated ewes during a superovulation protocol. It is well known that MAP serum concentration is not detected by the RIA kit used to measure P₄. For MAP detection, it is necessary to use a specific antiserum [30,31]. We speculate that this may occur because of the divergence in the chemical structure between MAP and P₄. The MAP molecule presents a methyl (CH₃) radical at C6 position and a methyl acetate (C₂H₃O₂⁻) radical at C17 position (Fig. 1), whereas in the P₄ molecule, this radical is absent. The exogenous P₄ is bioidentical to the endogenous (luteal) hormone [11].

The abundance of messenger RNA (mRNA) encoding proteins expressed in the oocyte (ie, MATER, ZAR1, BMP15, and GDF9), cumulus (ie, RELN, LRP8, FSHr, LHr, ERα, and STAR) or both (ie, BAX and Bcl-2) were evaluated in BCB⁺ COCs. In the P₄ group, 6 genes were upregulated (ZARI, GDF9, Bcl2, FSHr, LHr, and ERα), and in the MAP group, 2 genes were affected (RELN was downregulated and Bcl2 upregulated), both compared with the control (calibrator). However, comparing P₄ vs MAP (calibrator), 3 genes were upregulated (RELN, FSHr, and LHr) in the P₄ group.

The difference in RELN abundance (downregulated in MAP compared with control and upregulated in P₄ compared with MAP) was not related to serum P₄ concentration because RELN expression was no different between P₄ and control. The contrast in RELN mRNA expression may be due to the different chemical structure of the progestogens. We presume that P₄ and MAP pharmacokinetics activate distinct pathways, despite the similar therapeutic effect (pharmacodynamics). RELN expression is maximal in the theca cells of dominant follicles and binding LRP8 in granulosa cells by paracrine action, activating the downstream signaling pathways [32], which regulate the final follicle growth stage [7]. Yang et al. [33] reported RELN involvement with protein kinases activity and suggested it plays a role in progestogenic pathways. Conversely, Fayad et al. [32] have demonstrated that RELN and its ligand, LRP8, are predominantly expressed in the dominant follicles and similarly downregulated during the ovulatory process and luteinization. Thus, we considered the RELN abundance as an important marker of oocyte competence acquisition.

We associate the gene expression profile observed in the P₄ group to the rise of serum P₄ concentrations. Recently, Menchaca et al. [14] demonstrated that high serum P₄ induced by a P₄ intravaginal device enhances oocyte developmental competence by increasing the in vitro cleavage rate and embryo development in sheep (compared with nontreated ewes). Cuadro et al. [13] have reported similar effects in the in vivo embryo production system in sheep, which both did and did not receive exogenous P₄ during the growing phase of the follicular wave. The authors found an improvement in the percentage of fertilized oocytes after insemination, a greater number of transferable embryos per donor, and better quality of the collected embryos. Otherwise, when P₄ priming was compared with MAP treatment, Bartlewski et al. [15] reported no differences in terms of the number of corpora lutea and embryos recovered in superovulated ewes.

In support of the upregulation of ZARI in the P₄-treated group in relation to the control group, we considered this a good indication of quality improvement; moreover, it may corroborate the findings of Menchaca et al. [14] and Cuadro et al. [13], already mentioned. ZARI is a good predictor of developmental competence, mainly in immature oocytes. ZARI mRNA plays a role in early embryo development until the zygotic genome activation [34,35].

FSH levels directly influence FSHr, LHr, and ERα [8,36] by downstream activation of a FSHr/adenylylclc/ob/
PKA cascade to trigger estradiol (E₂) production [37–39]. An interaction between P₄ and FSH has already been evidenced [14,40], and we suggest that this interaction could be a function of the P₄ molecular structure; it may be that the methyl acetate radical on MAP molecules may prevent this interaction. Bartlewski et al [15] observed longer FSH-metabolic clearance rates in the P₄-treated than the MAP-treated ewes. Based on this finding, and those reported by Wei et al [36] and Knecht et al [41], we speculate that the longer FSH bioavailability in P₄-treated ewes possibly maintained the stimulus of downstream FSH-induced pathways, enabling the expression of these receptors, including LHR, through raising of cAMP, despite P₄ inhibition of LH release pulsatile frequency by feedback mechanism. The upregulation of these receptors (FSHr, LHR, and ERα) may promote better conditions for oocyte to respond to the in vitro maturation conditions regarding the role of hormonal regulation during the maturation process.

The greater serum P₄ also affected GDF9 expression, probably by extending FSH bioavailability in the P₄-treated group. Exogenous FSH affects GDF9, as we have previously demonstrated [8]. GDF9 stimulates granulosa cell proliferation and acts in the regulatory process during final follicle growth, before the LH surge before ovulation; cumulus cells require GDF9 to support the metabolic cascades and steroid production [42,43]. Hence, the P₄-treated group seems to promote more appropriate COC gene modulation.

Apoptosis proteins regulate cell death and are good markers of cell viability. The antiapoptotic Bcl2 gene was upregulated in both the P₄- and MAP-treated groups in the present study. Similar results were obtained by Thammsirir et al [3] in a short-term MAP-treated protocol. These evidence indicate the positive effect of exogenous progestagens applied during ovarian stimulation on the apoptosis modulation cascade.

Nilsen and Brinton [44] demonstrated the divergent impact of P₄ and MAP on the mitogen-activated protein kinase (MAPK) pathway for the mechanism of neuroprotection in neuron culture, which reinforces our hypothesis about both progestagens activating different downstream pathways, despite their similar therapeutic effects. Exogenous support of P₄ seems to exert a beneficial effect on oocyte quality, based on the profile of gene expression (compared with the MAP-treated and control groups). On the other hand, MAP had a negative effect, mainly because of the downregulation of RELN, since this extracellular matrix glycoprotein is involved in 47 distinct cell functions, including MAPK pathway [33], which is necessary for germinal vesicle breakdown and oocyte maturation [45]. However, to confirm this deleterious effect of MAP, future research is necessary to evaluate in vitro development and molecular features of the embryos, such as polyspermy regulation, cryopreservation, and/or implantation.

In conclusion, despite P₄ and MAP promoted obtaining COCs with similar quality related to morphological features and BCB test, they induced dissimilar effects on the molecular status of immature COCs in sheep. Exogenous P₄ appears to influence indirectly the oocyte competence through its positive impact on the COC gene expression (ZAR1, GDF9, Bcl2, FSHr, LHR, and ERα), probably because of its chemical structure (identical to the P₄ endogenous) and greater serum P₄ concentration, which induce this difference in its biological activity. However, the exact mechanism by which it occurs still needs to be elucidated.

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